# Original Research Respiration of Soil Enriched with Manure and Mineral Materials (Methodical Aspects)

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## Abstract

Methods of start (as a new one), reactive, basal, and long-term respiration determination in laboratory conditions were used for evaluating  $C_{org}$  transformation with  $CO_2$  release from sandy soil (Haplic Luvisol) enriched in field experiments with organic (manure) and mineral (clay, lime) materials. They proved to be useful in such experiments. The objective of the investigation was to find which treatments stimulate or inhibit  $C_{org}$  mineralization. Of all treatments used, the least susceptible to  $C_{org}$  mineralization in soil appeared to be the soil enriched with manure. The most susceptible was soil enriched with soil+manure+clay+lower dose of lime.

Keywords: manure, clay, lime, incubation, respiration, CO2

# Introduction

Soil respiration depends on many factors such as organic substrate, moisture, temperature, texture, and structure of soil, available nutrients, pH, heavy metal content, pesticides, and time of incubation, etc. [1-4].

To achieve optimal fertility it is necessary to maintain the correct proportion between the amount of organic substance and mineral components in soil. Various substances are introduced to soils to improve their fertility and increase crop production, including manure, clay, lime, and mineral fertilization [5-8]. Nearly all processes that regulate soil fertility are based mainly on the transformation of organic matter [2, 3, 9-11]. However, these processes have occurred in Poland too intensively because of the types of soils (mostly sandy), climatic conditions, and intensive agriculture that decreases humus resources [12, 13]. This is a harmful effect for agricultural ecosystems and for the environment (increase of  $CO_2$  evolution). organic carbon mineralization in soil is to measure the amount of  $CO_2$  released during incubation for a definite time and temperature [16-19].

To counteract this effect it has been proposed to create

The aim of this paper is to test various methods of the determination respiration activity of sandy soil (Haplic Luvisol) enriched in with organic and inorganic materials on the course and rate of organic carbon ( $C_{org}$ ) transformation. The objective of the investigation was also to find which treatments stimulate or inhibit  $C_{org}$  mineralization.

# **Materials and Methods**

# Field Experiment

For investigations soil samples were taken from six 2 m<sup>2</sup> plots from experimental plots with soil enriched with

organo-mineral complexes in soils that can stabilize organic substance levels [14] by enriching soil with organic and mineral matter resistant to decomposition [15]. One of the most frequently used methods to determine organic orthon mineralization in soil is to measure the

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Experimental plots – Treatments	Fertility substance, dose	
Ι	No treatment (untreated soil as control)	
II	Soil + manure (8 kg per plot)	
III	Soil + manure (8 kg per plot) + clay (50 kg per plot)	
IV	Soil + manure (8 kg per plot) + clay (50 kg per plot) + post-flotation lime (1.12 kg per plot)	
V	Soil + manure (8 kg per plot) + clay (50 kg per plot) + post-flotation lime (2.24 kg per plot)	
VI	Soil + manure (8 kg per plot) + clay (50 kg per plot) + $Ca(OH)_2$ (2 kg per plot)	

Table 1. Fertility substance and their doses on the plot.

All plots were covered with the grass Dactylis glomerata.

organic and inorganic materials (6 treatments, Table 1). Plots were located in Kolonia Boniewo near Lublin, Poland. They were established in 2001 on Haplic Luvisol sandy texture, pH in 1M KCl 4.46, with 0.46% of organic C. Haplic Luvisols in field culture are the majority in Poland.

Cattle manure (M) was taken from an agricultural farm and added to all soils except the control. M contains in dry mass: 2-2.5% N, 0.6% P, 2.8% K, and 0.8% Mg. Clay (Cl) came from the sulfur mine "Jeziórko" where it is used for soil improvement around the area of the mine [20]. Postflotation lime (L) was a waste product from sulfur processing.

# Laboratory Experiment

In the autumn of 2002 soil samples were taken (14 months from the addition to soil of the mentioned substances) from the upper horizons (0-25 cm) of the plots of field experiment to the laboratory, where 2.5 g of each soil portion (well homogenized in three replications) was placed in (60 ml) glass bottles, watered to 20% moisture content, and closed with rubber stoppers. The resulting soils were then incubated in three levels of time and temperature. The cumulative gases evolved from soils were collected with a syringe in amounts of 0.05 ml, and four methods of respiration measurements [21] were used.

The 1<sup>st</sup> incubation level was at 28°C for 24 h with  $CO_2$  collection after 5 min, 6 h, 12 h, and 24 h. The authors term this step start respiration (SR). It determines the use of dissolved organic C by microorganisms [21].

The 2<sup>nd</sup> level was at 28°C for seven successive days. The value of respiration after the first three days is named by Appfelthaler [16] reactive respiration (RR) and, after the next four days, basal respiration (BR). Gases were collected each day at a previously established fixed time (without washing flask by  $CO_2$ -free air).

The 3<sup>rd</sup> level at 25°C for five weeks with respiration measurements at one-week intervals was named according to Klimanek's [19] long-term respiration (LTR). Gases were collected at the end of each week.

Temperatures of 25°C and 28°C were selected as being optimal for the development of bacteria [1, 15].

## Measurement of CO<sub>2</sub>

Carbon dioxide was determined by a Shimadzu GC-14A gas chromatograph equipped with thermal conductivity detector (TCD) on a column filled with Porapak (TCD temperature 60°C, column temperature 40°C, gas flow 60 ml min<sup>-1</sup>) [21].

## Treatment of Results

 $CO_2$  data from each successive period of incubation were subtracted from those of the previous one and then divided by the incubation time expressed in minutes for SR and in days for RR, BR, and LTR. Figs. 1, 3, and 5 show cumulative amounts of  $CO_2$  released from soil.

The rate of  $CO_2$  release from soil during respiration is shown in Figs. 2, 4, and 6.

#### Statistical Analysis

The results were analyzed statistically using analysis of variation (Statgraphics; 95% LSD) and regression (Excel).



 $\label{eq:constraint} \mathsf{Treatments} \quad \Box \ \mathsf{I} \ \Box \ \mathsf{II} \ \blacksquare \ \mathsf{III} \ \blacksquare \ \mathsf{IV} \ \blacksquare \ \mathsf{V} \ \boxdot \ \mathsf{VI}$ 

Fig. 1. Cumulative amount of the first day of CO<sub>2</sub> release during start respiration (SR).

## Results

## Start Respiration

The cumulative amount of CO<sub>2</sub> released from soil after 5 min of SR incubation was in the range 4.21 to 5.53 CO<sub>2</sub>-C mg·kg<sup>-1</sup> (Fig. 1). In the subsequent hours of incubation, the respiration activity of enriched soils distinctly increased in relation to the control soil (from 1.2 to 2.3 times). After the first 24 h of incubation the highest rate of CO<sub>2</sub> production (52.7 mg CO<sub>2</sub>-C kg<sup>-1</sup>) was observed in treatment IV (soil +M+Cl+ 1.12 kg L), and the lowest (23.4 mg CO<sub>2</sub>-C kg<sup>-1</sup>) in the control soil. The data set was used for regression analysis of possible correlations between the amount of CO<sub>2</sub> release and time of incubation during particular types of respiration (SR, RR, BR, and LTR). During the first day of incubation no significant differences in an average value of CO<sub>2</sub> release between treatments were found.

Fig. 2 showes the rate of soil respiration (amount of  $CO_2$  per min) for the first day of incubation and shows that the highest respiration activity resulting from additives occurs during the first 5 min of incubation. Microbes in the control soil were the most active in the use of native  $C_{org}$  while the weakest carbon mineralization was found in treatment V (soil+M+CL+2.24 kg L). Clay and lime together with manure added to soil treatments III-V slowed this process only negligibly. In the next hours of incubation the rate of  $C_{org}$  mineralization was noticeably lower.

Respiratory activity dropped after 6 h of incubation, to increase again at 24 h.

In summary we can state that, in comparison to the control soil, the most intensive  $C_{org}$  mineralization during SR was in treatment IV (soil+M+Cl+ 1.12 kg L), and the weakest (not statistically significant) was in treatment VI (soil +M+L+ Ca(OH)<sub>2</sub>). The addition of M+Cl to soil stimulated the mineralization process, but the addition of clay, a higher dose of 2.24 kg L, and Ca(OH)<sub>2</sub> slowed it. The cumulative amount of  $CO_2$  released from soil during RR and BR types is shown in Fig. 3. The progressive increase (similar to SR) of the cumulative amount of  $CO_2$ release during soil incubation is clearly visible, during both RR and BR. Significantly higher cumulative evolution from soil during the first 3 days of incubation (RR) and the next 4 days (BR) was observed in treatments III (soil+M+Cl) and IV (soil+M+Cl+1.12 kg L).

During RR (Fig. 4) the highest respiration activity of soil was in the first day of incubation with a clearly visible effect of our additives. The rate of  $C_{org}$  mineralization dropped in the second day of incubation and increased slightly in the third day. High respiration activity was seen in treatments III and IV (soil+M+Cl and soil+ M+Cl+1.12 kg L) during the whole period of RR. Soil treated with other materials showed lower rates of  $C_{org}$  mineralization, close to that of the control soil. In all treatments the amount of mineralized  $C_{org}$  after three days of incubation was higher, ranging from 1.1 times in treatment VI (soil+M+Cl+1.12 kg L) compared to the control.

During BR (Fig. 4) the rate of  $CO_2$  production diminished considerably in comparison to RR, but toward the end of the experiment there was still a high daily rate of mineralization in treatments III (soil+M+Cl) and IV (soil+M+Cl+1.12 kg L). The lowest was in treatment II (soil+M). Therefore, nearly all materials used in field experiments stimulated the rate of  $CO_2$  production during BR. Additions of manure and lime distinctly prolonged stimulation of the respiratory process in soil toward the first day of soil incubation. However, similarly to SR, a dose of 2.24 kg lime and Ca(OH)<sub>2</sub> introduced to soil inhibited  $CO_2$  release in comparison to treatments III and IV (soil+M+L).



Treatments

Fig. 2. The rate of  $CO_2$  release from soil during start respiration (SR)



Treatments □ | □ || □ || □ |V ■ V ⊡ VI

Fig. 3. Cumulative amount of  $CO_2$  released from soil during the first 3 days (reactive respiration RR) and the following 4 days (basal respiration (BR) of soil incubation.



Treatments  $\Box | \Box | | \Box | | \Box | | \Box | V \Box V$ Fig. 4. The rate of CO<sub>2</sub> release from soil during reactive respiration (RR) and basal respiration (BR).

## Long-Term Respiration

The highest cumulative amount of  $CO_2$  release from soil (449 mg  $CO_2$ -C kg<sup>-1</sup>) was in treatment IV (soil + M+Cl+1.12 kg L) (Fig. 5). The lowest amount was found in the control soil (207 mg  $CO_2$ -C kg<sup>-1</sup>). Therefore, all materials used in the experiment stimulated  $CO_2$  release from soil during the whole period of  $C_{org}$  mineralization, including LTR. Differences between the mean values of  $CO_2$ release from soil treatments during LTR are evident.

The highest respiration activity of soil was observed during the first week of incubation and decreased systematically during the course of soil incubation (Fig. 6). This testifies to the varying susceptibility of  $C_{\rm org}$  to mineralization.

In all experimental treatments the amount of  $C_{org}$  mineralization after 5 weeks of soil incubation was 1.4 times higher in treatment VI (soil+M+Cl+ Ca(OH)<sub>2</sub>) and 2.2

times higher in treatment IV (soil+M+Cl+1.12 kg L) than in the control soil.

# Discussion

Respiration activity of soil is the most important characteristic of soil biological activity and depends on many factor [18], including carbon availability, temperature, and humidity. Our result showed that during the first 5 min of incubation (SR), microbes in the control soil were the most active in the use of native Corg, while the weakest carbon mineralization was found in treatment V (soil+M+Cl+ 2.24 kg L). In the next hours of incubation the rate of Corg mineralization was noticeably lower. This means that 6 h of incubation was sufficient for microbial adaptation to growth in experimental conditions and resulted in a higher respiration intensity of enriched soil in comparison to the control. Respiratory activity dropped after 6 h of incubation, to increase again at 24 h. These convertible cycles in the increase and decrease of CO2 evolution from soil were probably due to the exhaustion of a very easy oxidized C fraction and the consequent necessity of using C-bearing material more resistant to biological oxidation. These probably reflect differences in its availability as well as the adaptation of microbes to decomposition in subsequent C fractions. The smoothest dynamics of C<sub>org</sub> decomposition was observed in the control soil. The greatest fluctuations were in treatment V (soil+M+Cl+2.24 kg L). Therefore, we have concluded that the term start respiration is justifiable and can be useful in discussing the susceptibility of  $C_{org}$  to mineralization. The Corr content and its two fractions (easily available and not easy accessible) in the soil taken from the field experiment presented by Księżopolska [22] was very different (Table 2). A significant very close positive correlation between the rate of CO<sub>2</sub> release and easily available carbon for the SR (r=0.92\*\* for n=6) and negative correlation with not easy accessible fraction (r=0.868\*\* for



Treatments □ I □ II □ III □ IV ■ V ⊡ VI

Fig. 5. Cumulative amount of CO<sub>2</sub> release from soil during long-term respiration (LTR) of soil incubation.





Fig. 6. The rate of  $CO_2$  release from soil during long-term respiration (LTR).

Plots- treatments	Organic carbon	Easily available	Not easily available
	(g·kg <sup>-1</sup> )	$C_{org}$ (%)	$C_{org}$ (%)
Ι	6.0	35.0	65.0
II	7.5	20.2	79.8
III	8.2	18.4	81.6
IV	7.6	9.9	90.1
V	8.1	9.3	90.7
VI	7.8	9.6	90.4

Table 2. Total, easily available, and not easily available Corg [22].

n=6) was found. It confirmed our presumption that measuring SR determined the use of dissolved organic C by microorganisms.

During reactive and basal respiration significantly higher average value of CO<sub>2</sub> evolution from soil was observed in treatments III (soil+M+Cl) and IV (soil+M+Cl+1.12 kg L) in RR, and additionally in treatment V in BR. These additions significantly stimulated development of bacteria responsible for Corg mineralization compare to mineralization of native carbon in control soil. Much lower values were noted for remaining treatments and the lowest one for control soil. However, the addition of lime and hydrated calcium significantly decreased CO<sub>2</sub> release from soil in comparison to its treatment with manure and lime. This can be important in the storage of native and organic carbon introduced to soil. The stimulating effect of clay in Core mineralization found in our experiment appeared to be in contradiction to the inhibiting effect obtained by several other authors [6, 15]. They suggest this effect to be due to the formation of organo-mineral complexes resistant to carbon mineralization. In a laboratory such phenomena probably did not exist. However, Księżopolska [22] found slightly lower CO<sub>2</sub> emissions from plots with clay additions compared to plots without the addition under field experiment (the average value from three years lasted for the field experiment).

The addition of manure to soil slowed the rate of organic matter decomposition during BR in comparison to the control soil and to soils with other additives. The results obtained show that materials added to soil distinctly affected the availability of  $C_{org}$ . During the first day of incubation, microbes were the most active in the use of  $C_{org}$  introduced to soil of treatment II (soil+M). Similar results were obtained by Cerhanova et al. [17]. The addition of clay to soil only gradually slowed this process. The addition of lime to soil had no significant effect on the rate of  $C_{org}$  use.

During RR the rate of  $C_{org}$  mineralization was higher than during the consecutive four days (BR) of organic carbon. mineralization and the differences in carbon availability were found. In the 7<sup>th</sup> day of BR, respiratory activity was relatively high (compared to the previous three days) with Similarly to SR, the variability of time maxima and minima in  $CO_2$  evolution from soil during RR and BR indicates differing carbon availability and may also reflect adaptation of microbes to decomposition of consecutive carbon fractions. However, the dynamics of the processes in comparison to SR is different but more stable. In the case of RR and BR there wasn't any significant correlation between  $C_{org}$  content and  $CO_2$  release.

Lowering temperature as well as prolonging the incubation time (LTR) significantly affected differences between  $C_{org}$  mineralization in control soil and soil modified by addition of improving materials. In all enriched soils the amount of CO<sub>2</sub> released was significantly higher than in control soil. The effect of carbon mineralization delay observed during BR in treatments II and VI (soil+M and soil+M+Cl+Ca(OH)<sub>2</sub>) in relation to control soil was canceled during LTR through the time factor.

The greatest fluctuations in  $C_{org}$  mineralization rate during LTR was in treatment III (soil+M+Cl). This probably reflects the much differentiated availability of carbon introduced to soil in this treatment. The addition of lime to soil with manure and clay distinctly affected the stabilization of  $C_{org}$  mineralization rate at longer times and lower incubation temperatures in comparison to other types of respiration (SR, RR, and BR).

The rate of  $C_{org}$  mineralization in soil during LTR is more stable than during the SR, RR, and BR phases of incubation. The exceptions are treatments II and III (soil+M and soil+M+Cl). This means that reducing the incubation temperature by 3°C distinctly slows  $C_{org}$  mineralization and allows its available pool to be used for a longer time.

The most stable  $C_{org}$  mineralization rate in soil is observed in the control; this probably implies the best microorganism adaptation for  $C_{org}$  decomposition as well as the lower availability of carbon compounds subject to mineralization.

The highest and the most even equal respiration activity was found in the first 2 weeks of the experiment. From the 3<sup>rd</sup> week of incubation onwards, greater differences in the respiration activity of soil were observed, depending on the type of enrichment used. This was confirmed by regression analysis, which showed very close positive correlation between cumulative CO<sub>2</sub> release from the soil and total C<sub>org</sub> content (r=0.875\*\* for n=6), and not easily available C<sub>org</sub> (r=0.91\*\* for n=6).

Reversible cycles of the increase and decrease in the amount of  $CO_2$  released from soil seem to indicate the adaptation of different microorganisms to variations in  $C_{org}$  availability. Similar results were obtained by Klimanek [19], who found that during daily measurements of  $CO_2$  over 35 days of incubation, the highest soil respiration was during the first 10-15 days. It then diminished until the end of incubation.

# Conclusions

- Four methods of determining respiration activity of soil (start, reactive, basal, and long-term respirations) appeared to be useful in evaluating C<sub>org</sub> transformation with CO<sub>2</sub> release in sandy soil enriched with various organic and mineral materials.
- The amount of CO<sub>2</sub> released from soil in all experimental treatments and of respiration types was in the range of about 10 to about 27 mg –C day<sup>-1</sup>.
- 3. Of all treatments used, the least susceptible to C<sub>org</sub> mineralization in soil appeared to be soil enriched with manure, and the most susceptible was soil enriched with soil+manure+clay+lower dose of lime.
- 4. A significant positive correlation between the rate of  $CO_2$  release as easily available  $C_{org}$  and negative correlation with not easily available  $C_{org}$  for start respiration was found. Also, positive correlation appeared between mean values of  $CO_2$  release as total and not easily available  $C_{org}$  for long-term respiration.
- 5. The amount of  $CO_2$  release from soil enriched with manure, clay, and the lower dose of lime was more than two-fold higher in comparison to control soil, as well during basic and long-term respiration.
- 6. The highest intensity of CO<sub>2</sub> release from soil was found during start respiration and the lowest during long-term respiration.

# References

- FANG C., MONCRIEFF J.B. The dependence of soil efflux on temperature. Soil Biology and Biochemistry, 33, 155, 2001.
- GLIŃSKI J., STĘPNIEWSKI W. Soil aeration and its role for plants. CRC Press Inc., Boca Ration, Florida, 1985.
- HONG S.L., LING H.L., XING G.H., HUANG J.H., SUN J.X., WANG H.Y. Respiratory substrate availability plays a crucial role in the response of soil respiration to environmental factors. Applied Soil Ecology, 32, 284, 2006.
- LA SCALA N. Jr., PANOSSO A.R., PEREIRA G.T., GON-ZALEZ A.P. MIRANDA J.G.V. Fractal dimension and anisotropy of soil CO<sub>2</sub> emission in an agricultural field during fallow. International Agrophysics, 23, 353, 2009.
- HAYNES R.J., NAIDU R. Influence of lime, fertilizer and manure applications on soil organic matter content and soil physical conditions: a review. Nutrient Cycling in Agroecosystems, 51, 123, 1998.
- MALKAWI A.L.H., ALAWNECH A.S., ABU-SAFAGAH O.T. Effects of organic matter on the physical and the physicochemical properties of an illitic soil. Applied Clay Science, 14, 257, 1999.

- PREGITZER K., LOYA W., KUBISKE M. Soil respiration in northern forests exposed to elevated atmospheric carbon dioxide and ozone. Oecologia, 148, 503, 2006.
- KUBAT J., LIPAVSKY J. Steady state of the soil organic matter in the long-term field experiments. Plant, Soil, Environment, 52, 9, 2006.
- BAGGS E.M. Partitioning the components of soil respiration: a research challenge. Plant and Soil, 284, 1, 2006.
- VINCENT G., SHAHRIARI A.R., LUCOT E., BADOT P., EPROM D. Spatial and seasonal variations in soil respiration, in a temperate deciduous forest with fluctuating water table. Soil Biology and Biochemistry, 34, 2527, 2006.
- GAJDA A.M. Microbial activity and particulate organic matter content in soils with different tillage system use. International Agrophysics, 24, 129, 2010.
- KUSIŃSKA A., KISIEL M. Biodiversity effect on the processes of mineralization and humification of organic matter in a meadow ecosystem. International Scientific Conference "Humic Substances in Ecosystems 7" – Bachotek, Poland, 2007.
- TROJANOWSKI J. Transformation of soil organic matter. PWRiL, Warszawa, Poland, pp. 331, 1973 [In Polish].
- SCHARPENSEEL H.W. Preface to workshop measurement of carbon in tropical soils under global change: science, practice and policy. Geoderma, 79, 1, 1997.
- WANG A.J., DALAL R.C., MOODY P.W., SMITH C.J. Relationships of soil respiration to microbial biomass, substrate availability and clay content. Soil Biology and Biochemistry, 35, 273, 2003.
- APPFELTHALER R. Methods for the evaluation of the quantity and quality of soil organic matter and biological activity of soils. Res. Inst. Crop. Production: 19, 1994.
- CERHANOVA D., KUBAT J., NOVAKOVA J. Respiration activity of the soil samples from the long-term field experiments in Prague. Plant, Soil and Environment, 52, 21, 2006.
- FRESCHET G.T., MASSE D., HEIN E., SALL S., CHOTTE J.L. Long-term changes in organic matter and microbial properties resulting from manuring practices in an arid cultivated soil in Burkina Faso. Agriculture, Ecosystems and Environment, 123, 175, 2008.
- KLIMANEK E.M. Release of CO<sub>2</sub> from soil samples in a laboratory experiment. Agrobiological Research, 47, 380, 1994.
- GOŁDA T. The influence of reclamation of soil cultivation managements on sulphur transformation in post-flotation slime. Sozologia i Sozotechnika, 37, 1496, 68, 1993.
- WŁODARCZYK T., KSIĘŻOPOLSKA A., GLIŃSKI J. Some new method of soil respiration activity measuring. Teka Commission of Protection and Formation of Natural Environment, volume V<sub>A</sub>, 153, 2008.
- KSIĘŻOPOLSKA A. Effect of organic and mineral materials introduced to soil (Haplic Luvisol) on its properties and transformation of organic matter. Wydawnictwo Adam Marszałek, Toruń, Poland, pp. 154, 2010 [In Polish].